Relationship between Vitamin D Receptor Gene Polymorphism FokI and Fetuin-A as Marker of Vascular Calcification in Egyptian Hemodialysis Patients

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ABSTRACT

Background: Vascular calcification is a common complication in end-stage renal disease patients and the leading cause of morbidity and mortality.

Aim of the work: The current work aimed to evaluate the possible relationship between vitamin D receptor gene polymorphism FokI with fetuin-A and intact parathormone hormone in Egyptian hemodialysis patients.

Patient and Methods: This study is a cross-sectional study including 50 hemodialysis patients attending the hemodialysis unit in Al-Zahra University Hospital and 30 apparently healthy persons as a control group. For all studied subjects, a detailed history was taken, physical examination, carotid intima-media thickness by carotid duplex, echocardiography, Laboratory investigations included serum levels of calcium, phosphorus, iPTH, fetuin, and genotype frequency by PCR.

Results: There was a highly significant increase in carotid intimal media thickness in the patient group Vs. control group & a highly significant decrease in serum Fetuin-A in the patient's group Vs. control group. There was a significant difference in the genotype distribution of FokI polymorphism among patients and control. There were significant negative relationships between FF genotype and iPTH and a highly significant negative correlation between serum fetuin-A and iPTH in the patient group.

Conclusion: Vitamin D receptor gene FokI polymorphism FF genotyping is more frequent in hemodialysis patients who have higher iPTH and lower fetuin-A levels, which could identify the high-risk group susceptible to vascular calcification in hemodialysis patients. Further studies are needed.

Keywords: Vascular Calcification; Vitamin-D Receptor Polymorphism; FokI; Fetuin-A; Parathormone hormone.

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* Main subject and any subcategories have been classified according to the research topic.
INTRODUCTION

Cardiovascular disease is the primary cause of morbidity and mortality in chronic kidney disease [CKD]. Some studies reported that about 40% of hemodialysis patients [HD] developed cardiovascular diseases [CVD][1]. Vascular calcification is a prominent feature of arterial disease in CKD and may impact cardiovascular mortality by modulating both arteriosclerosis [arterial stiffening] and atherosclerosis[2]. The nature of vascular calcification is progressive and is associated with arterial stiffness and increased mortality. Age, duration of dialysis, and diabetes mellitus are clear determinants of the severity of calcification; however, novel insights into the pathomechanisms of unwanted calcification processes have been gained more recently. Disturbances of mineral metabolism such as hyperphosphatemia and hypercalcemia appear to contribute to progressive calcification, not only by passive precipitation but by actively inducing changes in vascular smooth muscle cell behavior toward an osteoblast-like phenotype. Specific calcium-regulatory proteins may act locally or systemically as calcification inhibitors. Dysregulation of them, including fetuin-A, matrix Gla protein, osteoprotegerin, and pyrophosphates, may also be pathophysiological relevant factors in the context of uremic extracellular calcification [3].

Increased serum level of parathormone hormone [PTH] and hyperphosphatemia are risk factors for CVD and increased HD patients’ mortality. Vitamin D signals through its receptors [vitamin D receptors] [VDR], a specific zinc-finger nuclear receptor. Vitamin-D functions are characterized as genomic, mediated through the VDR transcriptional effects inside the cell nucleus, and non-genomic when the VDR induces rapid signaling [4]. Genetic-association studies have demonstrated that variations in VDR function induced by polymorphisms at the 3’ and 5’ regions of the VDR gene may have an effect on mortality risk in hemodialysis patients [5]. The VDR is a member of the superfamily of nuclear hormone receptors. Its protein consists of 427 amino acids in humans, with a molecular mass of ~48 kDa[8]. A polymorphism is a genetic variant that appears in at least 1% of the population. It can occur in the coding or non-coding region of the genome. The majority of polymorphisms are silent, meaning that they occur in the non-coding region[introns] and therefore do not alter the function or expression of a gene. If a gene polymorphism is located in the coding region of the genome[exons], it can alter the expression of the gene and create variation within a given population. Because of their abundance in the human genome and their high frequency in the human population, it has often been studied to explaining variation in the risk for common diseases [7].

Several common genetic variants have been identified in the VDR gene, including the FokI[‘f’], BsmI[‘b’], Tru9I [‘u’], EcoRV, ApaI[‘a’], and TaqI[‘t’] polymorphisms, identified by a biallelic variation in a restriction endonuclease site and their name based on the restriction endonuclease[8]. The discovery of genetic variants linked with diseases’ susceptibility can be useful in preventive medicine. If a relationship with the disease emerges from association studies, this finding would strongly support the idea that the candidate gene is in some way involved in the disease [9].

AIM OF THE WORK

The current work aimed to evaluate the relationship between VDR gene FokI polymorphism and both fetuin-A level and iPTH as the main factors involved in vascular calcification in hemodialysis patients.

SUBJECT AND METHODS

The current work was across sectional study including 50 patients with end-stage renal disease on regular HD for at least 6 months, 4 hours/session, 3 times per week. Cases were selected from those attending hemodialysis units in Al-Zahra University Hospital from October 2016 to May 2017. Oral consent was obtained from all of them, approval for the study approval was obtained from Al- Azhar University’s ethics committee. They were 34 males [68%] and 16 females [32%], their age ranged from 28 to 77 years, thirty apparently healthy subjects were served as the control group, they were17 males [56.7 %] and 13 females [43.3%], their age ranged from 35 to 70 years. None of the patients had vitamin D therapy or therapy for high PTH. Ischemic heart diseases, liver cirrhosis, infection, and inflammatory bowel disease were excluded.
All Patients and control groups were subjected to; careful history and full clinical examination. Echocardiography, Doppler study for carotid intima-media thickness [CIMT] Laboratory investigations showed that 2samples were taken from the patients. 5ml whole blood on EDTA for genotype frequency in which Genomic DNA was extracted from whole blood samples using the QIaAmp DNA Blood Midi Kit. Genotyping of the FokI [rs2228570] polymorphism was performed using the polymerase chain reaction fragment length polymorphism [PCR-RFLP] analysis. The FokI polymorphism was detected using forward primer 5-AGC TGG CCC TGG CAC TGC TCT-3 and reverse primer 5-ATG GAA ACA CCT TGC TTC TTC TCC-3, which amplify 265 base pair [bp] fragment containing FokI site. PCR was performed for the FokI polymorphism at the following condition:

After denaturation at 95 °C for 5 min, 35 cycles were performed with denaturation for 45 s at 94 °C, hybridization For 45 s at 60 °C, and elongation for 45 s at 72 °C; the last cycle was followed by an extension step of 10 min at 72 °C. PCR products were checked by electrophoresis on 2% agarose gel. The PCR products were digested with the respective restriction Enzymes according to the manufacturer’s instructions as follows: at 37 °C for 5 min with FokI Digested products separated in 3% agarose gels and visualized by ethidium bromide staining and genotype were determined according to the digestion pattern. Furthermore, FokI polymorphism is confirmed by PCR repetition and PCR product commercial sequencing [genfanavaran]. Digested products for FokI polymorphism contain 196 bp and 69 bp530 bp and 210 bp fragments that indicated f allele and undigested alleles as F. And 5ml serum for fetuin A and other tests which include fasting Blood sugar [FBS], blood urea, serum creatinine, Serum Calcium, Phosphorus, iPTH, Serum Albumin, total cholesterol and triglyceride. As regard to Fetuin A; Pre-dialysis blood sampling from a peripheral vein was performed after 12 h of overnight fasting. Subsequent serums were separated within 30 min and samples were kept frozen at −70 °C until analysis was done. In the BioVendor Human Fetuin-A ELISA, Standards, Quality Controls and samples are incubated in microplate wells pre-coated with polyclonal anti-human Fetuin-A antibody. After 60 minutes’ incubation and washing, polyclonal anti-human Fetuin-A antibody, conjugated with horseradish peroxidase [HRP] is added to the wells and incubated for 60 minutes with captured Fetuin A. Following another washing step, the remaining HRP conjugate is allowed to react with the substrate solution [TMB]. The reaction is stopped by the addition of an acidic solution

The absorbance of the resulting yellow product is measured spectrophotometrically at 450 nm. The absorbance is proportional to the concentration of Fetuin-A. A standard curve is constructed by plotting absorbance values against concentrations of Standards, and concentrations of unknown samples are determined using this standard curve. Most microplate readers perform automatic calculations of analyte concentration. The standard curve is constructed by plotting the mean absorbance at 450 nm [Y] of standards against the log of the known concentration [X] of standards, using the four-parameter algorithm, and results are reported as the concentration of Fetuin-A [ng/ml] in samples. The complete blood picture was determined on Coulter Counter T890 was used [Coulter Counter, Hardened, UK]. Determination of fasting blood sugar, kidney function tests, and lipid profile were done on Hitachi auto analyzer 736. Calcium and phosphorus were determined using standard laboratory techniques with commercial kits]. Serum Albumin was assayed by an enzymatic colorimetric method with an automated chemical analyzer. Serum iPTH was determined by enzyme-linked immunosorbent assay [ELISA] using the Immunodiagnostic system.

**Statistical analysis:** Statistical analysis was conducted, tabulated, and analyzed using Statistical Package for Social Science [SPSS] program, software version 20. All continuous variables were expressed as mean ± standard deviation. The data were analyzed using the Student and chi-square test. Calculation of Spearman coefficient to determine the correlation between biochemical parameters. P values less than 0.05 were considered significant, and more than 0.05 none significant.
RESULTS

As regard age and sex, there was no significant difference between the studied groups. The causes of ESRD were: 31 patients [62%] were hypertensive, 11 patients [22%] with diabetic nephropathy, 4 patients [8%] of unknown etiology, 1 patient [2%] with polycystic disease, 1 [2%] amyloidosis, 1 [2%] lupus nephritis, and 1 patient [2%] with analgesic nephropathy [Table 1]. There was a highly significant increase in CIMT in the patients’ group than in the control group [Figure 1]. Echocardiographic findings in the patient group were 23 patients [46%] with the calcific aortic valve, 12 with the calcific mitral valve, and 3 [6%] with both Calcific aortic and mitral valve, and 12 [24%] with normal echocardiographic finding [Table 2]. There was a highly significant decrease in serum Fetuin-A in patient’s group [36.5 ± 8.05] ng/ml in comparison to control group [82.5 ± 9.12] ng/ml [P-value < 0.001] [Figure 2]. There was a statistically significant negative correlation between serum fetuin A and iPTH in the patient group [r = -0.851 P= 0.000] [Figure 3]. There was no significant correlation between valvular calcification and fetuin-A in hemodialysis patients [Table 3]. There was a highly significant difference in the genotype distribution of FokI polymorphism among patient and control groups P-values 0.001. In patient group, the distribution frequencies of FF, Ff, and ff genotypes were 32 [64%], 18 [36%] and 0 [0%] respectively, while in control group the distribution frequencies of FF, Ff, and ff Genotypes were 16 [53.3%], 0 [0%] and 14 [46.7%] respectively [Table 4]. There was a significant increase in iPTH in FF genotype than Ff genotype p value = 0.014, while there was no significant relation with other lab parameters [Figure 4]. There was no significant relation between fokI polymorphism genotype frequencies and fetuin-A in the patient group.

Table [1]: Baseline characteristics of studied groups

<table>
<thead>
<tr>
<th>Patients group</th>
<th>Control group</th>
<th>Test value</th>
<th>Pvalue</th>
<th>Sig.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age.</td>
<td>Mean± SD</td>
<td>Range</td>
<td></td>
<td></td>
</tr>
<tr>
<td>No. = 50</td>
<td>52.06 ± 12.68</td>
<td>28 – 77</td>
<td>0.779*</td>
<td>0.438</td>
</tr>
<tr>
<td>No. = 30</td>
<td>54.17 ± 9.85</td>
<td>35 – 70</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Males</td>
<td>34 [68.0%]</td>
<td>17 [56.7%]</td>
<td>1.042*</td>
<td>0.307</td>
</tr>
<tr>
<td>Female</td>
<td>16 [32.0%]</td>
<td>13 [43.3. %]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>The underlying kidney disease</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hypertension</td>
<td>31</td>
<td>62.0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Diabetic nephropathy</td>
<td>11</td>
<td>22.0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Unknown etiology</td>
<td>4</td>
<td>8.0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Polycystic kidney disease</td>
<td>1</td>
<td>2.0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amyloidosis</td>
<td>1</td>
<td>2.0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lupus nephritis</td>
<td>1</td>
<td>2.0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Analgesic nephropathy</td>
<td>1</td>
<td>2.0%</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Figure [1]: Comparison between patients and control group as regard carotid intimal media thickness
Table [2] Echocardiographic finding in the patient group

<table>
<thead>
<tr>
<th>Echocardiographic finding</th>
<th>Patients group [N=50]</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No.</td>
</tr>
<tr>
<td>Normal echo</td>
<td>12</td>
</tr>
<tr>
<td>Calcific aortic valve</td>
<td>23</td>
</tr>
<tr>
<td>Calcific mitral valve</td>
<td>12</td>
</tr>
<tr>
<td>Calcific aortic and mitral valve</td>
<td>3</td>
</tr>
</tbody>
</table>

Figure [2]: Comparison between patients and control group as regard fetuin A

Figure [3]: Correlation between serum fetuin-A and iPTH of the patient group. [iPTH: intact parathormone hormone]
Table [3]: Relation between serum fetuin-A and echocardiographic finding of patients group:

<table>
<thead>
<tr>
<th>Fetuin A</th>
<th>Test value</th>
<th>P-value</th>
<th>Sig.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean ± SD</td>
<td>Range</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Normal echo</td>
<td>38.17 ± 5.59</td>
<td>29 – 48</td>
<td>1.172</td>
</tr>
<tr>
<td>Calcific aortic valve</td>
<td>34.78 ± 8.14</td>
<td>22 – 52</td>
<td></td>
</tr>
<tr>
<td>Calcific mitral valve</td>
<td>39.08 ± 9.90</td>
<td>25 – 52</td>
<td></td>
</tr>
<tr>
<td>Calcific aortic and mitral valve</td>
<td>32.67 ± 5.86</td>
<td>26 – 37</td>
<td></td>
</tr>
</tbody>
</table>

NS: Non significant; P ≤0.05 Significant

Table [4]: Distribution frequencies of FokI polymorphism genotype among studied groups:

<table>
<thead>
<tr>
<th>Vit .D receptor Genotype</th>
<th>Patients group</th>
<th>Control group</th>
<th>Test value</th>
<th>P-value</th>
<th>Sig.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ff</td>
<td>No. = 50</td>
<td>0</td>
<td>0.0%</td>
<td>14</td>
<td>46.7%</td>
</tr>
<tr>
<td>FF</td>
<td>32</td>
<td>64.0%</td>
<td>16</td>
<td>53.3%</td>
<td></td>
</tr>
</tbody>
</table>

HS: Highly significant. P ≤0.001: Highly Significant

Figure [4]: Relation between FokI polymorphism genotype frequencies and iPTH of the studied group.

iPTH: intact parathormone hormone

DISCUSSION

Vascular calcification is commonly observed in patients with chronic kidney disease, and accumulating evidence indicates that its prevalence and severity correlate with long-term outcomes [10]. It was estimated that the age-adjusted cardiovascular events rate increases from 2.11 events per 100 person-years for individuals with normal GFR to 3.65 for those with CKD stage 1, to 36.60 for individuals with ESRD. Indeed, at all CKD stages, CVD mortality is disproportionately high compared to the general population. [11].

Our study detects the possible relationship between vitamin D receptor gene polymorphism FokI with fetuin-A and iPTH in Egyptian hemodialysis patients. There was no significant difference between the studied groups regarding age and sex, which means that both groups were comparable. In the current study, there was a highly significant increase in CIMT in the patient group than the control group; this agrees with the study done in Alexandria by El-Attar et al. [12], who compare 70 Egyptian patients on regular HD and 30 healthy subjects and found a significant increase in CIMT in hemodialysis patients.
compared to the control group and Paul et al.\textsuperscript{[13]}, who found that hemodialysis patients had significantly greater CIMT than age and sex-matched non-dialyzed chronic kidney disease (CKD) patients, suggesting that hemodialysis is an independent risk factor for atherosclerosis in CKD.

In a recent study done on HD patients with and without cardiovascular disease, the authors concluded that subjects with CVD have higher CIMT than those without CVD and men had higher CIMT than women, age was the most important marker for CIMT in patients on maintenance HD\textsuperscript{[14]}. Calciﬁcation of the aortic and mitral valves is a common finding in HD patients with a prevalence of four to ﬁve times higher than that in the general population. In a study done in Morocco on hemodialysis patients, the authors concluded that the prevalence of valvular calcification was 15\% with aortic valve location and mitral valve location in 41.2\%, and only hemodialysis duration seems to be associated with the occurrence of calcifications and approaches the marginal level of significance\textsuperscript{[15]}. In our study, cardiac valves calciﬁcation followed aortic valve 46\%, the mitral valve 24\%, and both aortic and mitral valve 5\%. The difference might be attributed to many factors such as differences in the study populations, mean age and dialysis vintage, different deﬁnitions and diagnostic methods for detecting CVC, the type of phosphate binders used, and dialysate calcium concentration. In another study done by Lin and his colleagues on HD patients with a median follow-up duration was 66 months, de novo cardiac valves calcifications developed in 45.98\% subjects: 58 developed aortic valve calciﬁcation alone, 42 developed calciﬁcation on mitral valve alone, and 20 developed both aortic and mitral valve calciﬁcation\textsuperscript{[16]}.

Sayarlioglu et al. studied 129 patients on hemodialysis, and their results showed a 23.3\% prevalence of mitral valve calciﬁcation, 21.7\% of aortic valve calciﬁcation, and the overall 33.3\% prevalence of valvular abnormalities\textsuperscript{[17]}.

In a recent Iranian study, the prevalence of mitral calciﬁcation, aortic and mitral annulus calciﬁcation, 57\%, 54\%, and 55\% respectively, and the overall valvular calciﬁcation was 73\% is higher than the rates reported by other studies\textsuperscript{[18]}.

Fetuin-A is produced in the liver and acts as a strong inhibitor of calcium - phosphate deposition. It can regulate several of the key cellular events that lead to VSMC calcification, including apoptosis and phagocytosis. It interacts directly with matrix vesicle release and forms stable colloid spheres with calcium and phosphorus, so-called “calciprotein particles” that inhibit hydroxyapatite precipitation and modulate vascular calciﬁcation processes locally and at early stages\textsuperscript{[19]}. When we compared serum fetuin A level in hemodialysis patients and the control group, we found that fetuin-A is signiﬁcantly decreased in patients than the control group; these in agreement with a recent study, which found serum fetuin A level was signiﬁcantly decreased in HD patients while osteopontin was signiﬁcantly higher in the patient group and no signiﬁcant correlation was found between patients with VC and patients without VC in terms of fetuin-A, osteopontin, and 25-OH-vitamin D levels and the authors concluded that VC is a frequent sign in patients undergoing HD and is not related to serum fetuin-A and osteopontin\textsuperscript{[20]}. In contrast to our results, Herman et al., in the cross-sectional study, reported that Fetuin-A was not different between dialysis patients and healthy subjects and could not be identiﬁed as an independent predictor of aortic stiffness\textsuperscript{[24]}. There was a signiﬁcant negative correlation between serum fetuin-A and iPTH in the patient group; these in agreement with Amani et al.\textsuperscript{[23]}. In contrast to our results, Shahnam-Valizadeh et al. found no correlation between Fetuin-A and iPTH in HD\textsuperscript{[21]}. There was no signiﬁcant correlation between valvular calciﬁcation and fetuin A in hemodialysis patients in the present study. These in agreement with Ossareh et al., who concluded that, the Fetuin-
A level was not different between patients with and without CVC and was not recognized as a predictor, while age, calcium level, and diabetes were identified as the most important predictors of calcification in their patients [18]. The BsmI and/or FokI VDR polymorphisms have been recognized as risk factors of some autoimmune diseases, including rheumatoid arthritis, Behçet's, graves' and Addison's diseases, multiple sclerosis, type 1 diabetes, and others. Moreover, both polymorphisms have been determined to be risk factors for colorectal, breast, prostate, and other cancers. The FokI and BsmI polymorphisms of the vitamin D receptor gene are regarded as strong markers of disturbed vitamin D signaling pathway so that VDR polymorphisms could be involved in secondary hyperparathyroidism chronic renal failure [25].

In our study, the distribution frequencies of FokI polymorphism in HD patients were 64% had FF genotype, 36% had Ff, and 0% were ff genotype while the FF genotype was in the control group 53.3%, Ff was 0%, and ff were 46.7% which was highly significant. In an Egyptian study done by El-Attar et al., they found the distribution frequencies of FokI polymorphism were FF, Ff, and ff genotypes in patient group 57.1 %, 31.4 %, and 11.4 %, respectively; while in the control group were 56.7 %, 40 %, and 3.3 % respectively and they concluded that there was no significant difference in the genotype distribution of FokI polymorphism among the control group and the total patients' group [12]. In another study, the prevalence of genotypes for FokI polymorphism was 32.6% FF, 39.1% Ff, and 28.3% ff, in Iranian HD [21].

The difference in our and other study results could result from the influence of geographical location and ethnic differences or study sample size related to the distribution of the VDR gene polymorphisms.

There was a significant increase in iPTH in the patient group's FF genotype in our study. In agreement with our results, the study of Vigo et al. found that iPTH level in the FF group was significantly higher than both Ff and ff groups in Spanish patients with CKD, and they suggested that FokI polymorphisms of the VDR gene may determine the parathyroid response in CRF patients[26]. In contrast to our results, El-Attar et al. did not found a significant difference in genotype distribution of FokI polymorphism among patients with iPTH less than 300 pg/ml and patients with iPTH more than 300 pg/ml [12].

In the present study, there was no significant relation between FokI polymorphism and serum fetuin-A in HD patients; these agree with Shahnan-Valizadeh etal[21].

**Conclusion:** Vitamin D receptor gene FokI polymorphism FF genotyping is more frequent in hemodialysis patients who have higher iPTH and lower fetuin A levels, which could identify the high-risk group susceptible to vascular calcification in hemodialysis patients and affect the management of cardiovascular events.

**REFERENCES**


